In the Specification:

Please amend the specification as shown:

Please delete paragraph [00134] and replace it with the following paragraph:

[00134] Suitable DNA constructs were designed and made as follows. Sequence GGTACCAAACTAGT **(SEQ ID NO: 1)** containing Kpn I and Spe I sites was inserted in the site coding for Gly92 and Val93 within the rat αo DNA (carried in a PQE6 vector) using QuikChange (Stratagene, 11011 N. Torrey Pines Road, LaJolla, CA 92037). A PCR (polymerase chain reaction) derived CFP fragment with the same sites was introduced and the αo-CFP was transferred into pENTR11 vector (Invitrogen, 1600 Faraday Avenue, P.O. Box 6482, Carlsbad, CA 92009). A PCR derived YFP fragment was made with a blunt-ended 5'-terminus and a Kpn I site at the 3'-end with the stop codon removed. A PCR fragment encoding the bovine ß1 subunit was made with a Kpn I site at the 5'-end and an EcoR I site at the 3'-end. Both fragments were sequentially inserted into the pENTR11 vector. A bovine y5 fragment was inserted between EcoRI and XhoI sites of pENTR11. This construct was cut with Xmn I and Sal I, end filled with Klenow polymerase and relegated to remove a Nco I site from the pENTR11 polylinker. Where necessary, the nucleic acid sequence of the DNAs were determined and/or their identity was checked and confirmed by restriction endonuclease digestion.

Please delete paragraph [00215] and replace it with the following paragraph:

[00215] *Fluorescence spectroscopy of insect cells expressing α-CFP tethered to the M2 receptor and βy-YFP*: Sense primer, AAAGTCGAACATGGGATGTACTCTGAGCGCAGAGGAGAGAGCCGCC **(SEQ ID NO: 2)** and anti-sense primer, TTTAGATCTTCAGTACAAGCCACAGCCCCGGAGATTGTTGGC **(SEQ ID NO: 3)**, were used to PCR amplify the α-o-CFP DNA. The product was digested with SalI and Bg1II and ligated to the M2 cDNA in the plasmid, pEntr1A, which was cut with the same enzymes downstream of M2. The construct was transferred into the pDest8

vector from Invitrogen using the LR Clonase enzyme. Baculoviruses were generated using standard methods. Insect cells (Sf9) were infected with M2-αo-CFP, β1 and YFP-γ5-wt viruses at a density of $1.0x10^6$ cells/ml. The cells were centrifuged and dissolved in PBS buffer at a density of $2.0x10^6$ cells/ml and measured in 250 μL cells in the Fluoromax 3 fluorometer. The excitations wavelength was set to 433 nm and emission spectrum was recorded from 460-580 nm. The excitation and emission slit-widths were 5nm, integration time was 0.5 sec/wavelength and monochromator increment was 2nm. First the spectrum was recorded. Then 2.5 μl of 10 mM carbachol (final conc. is 100 μM) was added and the cells were mixed up and down and the second spectrum was recorded. Untransfected cells were also measured under same conditions and untransfected cell spectrum was subtracted from these spectra.